



Indian Journal of Agriculture and Allied Sciences

A Refereed Research Journal

ISSN 2395-1109

e-ISSN 2455-9709

Volume: 4, No.: 2, Year: 2018

www.ijaas.org.in

Received: 10.06.2018, Accepted: 25.06.2018

Publication Date: 30th June 2018

INTEGRATED DISEASE MANAGEMENT IN SOLANACEOUS VEGETABLES

Vikas Gupta¹, Dorjay Namgyal¹, Anil Kumar², Phuntsog Tundup¹, Deldan Namgyal¹,
Rigzin Safal¹ and Shubana Bhat³

¹Krishi Vigyan Kendra-Leh (Ladakh), SKUAST-Kashmir, ²High Mountain Arid Agriculture Research Institute-Leh, SKUAST-Kashmir and ³Division of Plant Pathology, SKUAST-Kashmir, E-mail: vgskuastpathology@gmail.com, Corresponding Author: Vikas Gupta

Abstract: The losses due to the attack of insects and diseases to food and fruit crops are enormously large in terms of quantity, quality and monetary value. Solanaceous vegetables are severely attacked by the diseases and pests because solanaceous vegetables have fruits which are pulpy and succulent in nature, thus easily attracted by the pests and insects. Chemical Pesticides has led to many problems, such as pests and pathogens developing resistance, resurgence of once minor pest and pathogens into a major problem, besides environmental and food safety hazards. Integrated pest management (IPM) is smart and intelligent approach, which combines biological, cultural, physical and chemical control strategies in a holistic way for managing the diseases and pests in eco-friendly way without any hazard on human health and environmental resources.

Keywords: Disease, Integrated, Solanaceous and Vegetables.

Introduction: Plants are not absolutely safe in nature because of their susceptibility to various biotic factors like pests and diseases. These biotic factors constitute important constraints for economically feasible cultivation of various crops. The effect of damage and loss due to the attack of insects and pathogens is enormous, and in many cases resulting in complete loss in terms of crop yield. There is a strong historical background to support the fact that insects and diseases caused complete elimination of crops and farmers out of duress had to migrate to other places to escape from the menace of pests and put in renewed effort for cultivation of crop with the idea that such a shifting of place could help them to overcome the problem. The classic examples of insect and diseases attack can be highlighted by giving some examples like Irish famine in Europe due to late blight of potato in 1940s, coffee rust in Sri Lanka towards the end of 19th century, Bengal famine due to brown spot disease of rice crop in 1943, citrus decline in orange plantations, etc. Incidences of *Pyrilla* and top borer in sugarcane, pink boll worm in cotton, gall midge and brown plant hopper in rice, codling moth in apple and apricot in Ladakh

region are some of the examples of diseases and insects invasion. The losses due to the attack of insects and diseases to food and fruit crops are enormously large in terms of quantity, quality and monetary value. Though the disease and pest management schedules and practices are well documented and followed in most of the crops but, quite often the indiscriminate and unscientific use of pesticides has led to many problems, such as pests and pathogens developing resistance, resurgence of once minor pest and pathogens into a major problem, besides environmental and food safety hazards. Integrated pest management has become an alternate and most important of plant pest management practices, as chemical pesticides possess adverse effect on environment and human health. Integrated pest management (IPM), which combines biological, cultural, physical and chemical control strategies in a holistic way is an ideal approach of managing the diseases and pests. Rather than being dependent upon using a single component integrating all the available and feasible are more beneficial, effective and sustainable. IPM refers the dynamic and broad multidisciplinary approach biological,

cultural, and other alternatives, including safe and judicious use of pesticides. Its prime and foremost objective is to maintain pest levels below economically damaging levels while minimizing harmful effects of pest management on human health and environmental resources and maximizing the outputs of the produce.

Plants belonging to the group of Solanaceous crops include some of the very important vegetables like tomato, potato, chilli, capsicum, etc. Potato being an exceptional case as it is a tuber crop which has underground growth. Solanaceous crops are severely attacked by the diseases and pests. Commonly solanaceous vegetables have fruits which are pulpy and succulent in nature, thus easily attracted by the pests and insects. Insects directly feed upon these crops or sometimes act as vectors for various pathogens such as viruses.

Major Causal Organisms Associated with Solanaceous Vegetables

i) Fungi: A number of fungal pathogens have been found associated with the diseases of solanaceous vegetable in different parts of the world. *Pythium aphanidermatum* (Edson) Fitz. is a notorious soil borne pathogen causing pre- and post-emergence damping-off of seedlings and root and fruit rots of many crops^[1,2]. Seed rot and damping off have been reported to be incited by *P. aphanidermatum*, *P. adhaerens* Sparrow, *P. acanthircum* Drech., *P. angustatum* Sparrow, *P. irregularae* Buisman, *P. paroecandrum* Drech, *P. rostratum* Butler, *P. splendens* Brawn, *P. vexans* Debary and *P. ultimum* Trow^[3]. Reported that *P. aphanidermatum* was most virulent among the *Pythium* species^[4]. *Pythium* species have been reported as parasites of various economically important crop plants of solanaceous family, causing seed, seedling or root rots and damping-off diseases^[5]. Isolated *P. aphanidermatum* from brinjal fruit left for seed^[6], damped off and seedlings rot in chilli fruits and damped off in tomato seedlings. Besides, *P. aphanidermatum*, *P. ultimum* and *P. arrhenomanes* were commonly found associated with the damping-off disease of different host plants, including vegetables^[7].

Rhizoctonia solani (teleomorph: *Thanatephorus cucumeris*) is also a very common soil-borne pathogen, with worldwide distribution and a great diversity of host plants including bean, alfalfa, peanut, soybean, cucumber, papaya, eggplant, corn and many more^[8]. It is one of the most important causal organisms responsible for damping-off in

vegetable crops. Reported *R. solani* as causal organism of crown root rot of capsicum^[9], characterized by defoliation and wilting, resulting in the death of the plant. *R. solani* was one of the predominant fungi isolated from roots and stems of infected plants of bell pepper showing symptoms of wilting^[10]. Isolated and identified *R. solani* as causal agent of root rot and basal canker in egg plant^[11].

Fusarium species are known to cause wilt and damping off in several solanaceous vegetables. Isolated and identified *F. solani*, *Alternaria alternata*, *Coeliobolus lunatus*, *Cladosporium fulvum*, *Helminthosporium spiciferum* and *Tricothecium roseum* from diseased plants and fruits of aubergines^[12]. Observed high incidence of wilt at fruiting stage and identified *F. solani* as the casual agent of wilt^[13]. Identify *Fusarium oxysporum* f. sp. *melongenae* as the casual agent of Fusarium wilt on aubergine^[14]. Isolated *F. oxysporum* from the discoloured vascular tissue of brinjal^[15] and the isolate was tested for pathogenicity and confirmed as *F. oxysporum* f. sp. *melongenae*. Isolate *F. equisetii* from damped-off seedlings of brinjal^[16]. Reported that Fusarial wilts in solanaceous crops were caused by *F. oxysporum* f. sp. *lycopersici* (tomato), *F. oxysporum* f. sp. *melongenae* (brinjal), *F. oxysporum* f. sp. *vasinfectum* (pepper), *F. oxysporum* (chilli) and *Fusarium* spp. (potato)^[17]. Isolated *F. oxysporum* f. sp. *melongenae* from the brinjal plant roots showing wilt symptoms^[18].

ii) Bacteria: The bacterial wilt in solanaceous vegetables known to be caused by *Pseudomonas solanacearum* (Smith), limits the vegetable cultivation in humid, tropical, sub-tropical and warm temperate areas of the world, where the disease represents a major constraint in the production of numerous agricultural crops^[19]. The disease has been reported from different parts of the world, in U.S.A, bacterial wilt caused by *P. solanacearum* has been reported to be common on peppers, tomato, potato, eggplant and a number of other cultivated as well as wild plants^[20]. In India,^[21] reported that chilli could be one of the hosts for the strains of *P. solanacearum*, isolated from eggplants and potato. It has also been reported to attack variety of other crop plants like potato, tomato, brinjal, chilli, castor, groundnut, banana, etc.^[22, 23]. Reported *Ralstonia solanacearum* (syn: *Pseudomonas solanacearum*), the causal organism of chilli wilt^[24], as the most important constraint in chilli production in Kerala. Have

also observed that bacterial wilt was caused by *R. solanacearum* in tomato, chilli, tobacco, banana, brinjal, marigold, jute and ginger in West Bengal (India) [25].

Integrated Disease Management: The four basic requirements for management of plant diseases are clean and healthy seed, clean field or pathogen free soil, prevention of entry of infection of a pathogen in a standing crop and precaution during harvesting and storage of the produce. An ideal schedule for managing a disease is to integrate measures covering these four requirements. Integrated disease management attempts to use all the known suitable techniques of control to maintain a particular pest population at a level below that which causes economically important losses to the crop.

Reported that damping-off of vegetables (*Pythium* spp., *Fusarium* spp., *Rhizoctonia solani* and *Phytophthora* spp.) [26] was controlled by sowing hot water treated seed (52°C for 30 min) in 2 per cent formalin treated soil. Reported that integration of metalaxyl with *Trichoderma harzianum* had an additive effect on *Pythium* disease control in sugar beet [27]. Metalaxyl (0.01 %) provided only 62 per cent control but addition of 3.5 g/m of *T. harzianum* inoculum which was ineffective alone increased disease control up to 92.5 per cent along with metalaxyl. While higher amount of *T. harzianum* inoculum (10.5 g/m) in combination with either 0.01 or 0.05 per cent metalaxyl resulted in 100 per cent control of damping-off (*Pythium* spp.) disease of sugar beet. Reported that integration of soil solarization and seed treatment with Apron, *T. harzianum* and *Pseudomonas fluorescens* effectively reduced the population of damping off pathogens (*Pythium* spp., *Fusarium* spp., *Rhizoctonia solani* and *Sclerotium rolfsii*) in vegetable crops [28]. Used farm yard manure (10 t/ha), poultry manure (1.4 t/ha), mushroom spent compost (10 g/L of water), carbendazim (0.2 per cent) and copper oxychloride (0.3 per cent) against *Ralstonia solanacearum* and *Fusarium oxysporum* f. sp. *Melongenae* [29]. They stated that mushroom spent compost + paddy straw, *Trichoderma viride* and copper oxychloride effectively reduced the incidence of brinjal wilt.

Reported that pre-sowing seed treatment with *Pseudomonas fluorescens* + carboxin resulted in 62 per cent wilt control in lentil (*F. oxysporum* f. sp. *lentis*) in pots while seed treatment with carbendazim + thiram and *Gliocladium virens* + *P. fluorescens* + carboxin

were effective in controlling 48.8 and 44.2 per cent lentil wilt, respectively, in field [30]. Evaluated the efficacy of chemicals (salicylic acid, Bactrimycin and Plastomycin) [31], soil amendments (bleaching powder + lime + oil cakes and neem cake) and biocontrol agents (*P. fluorescens* MPf-1 and MPf-2, and *Trichoderma* sp.) against *R. solanacearum* causing wilt in tomato, and found that reduction in wilt incidence was highest with MPf-2 treatment followed by soil amended with bleaching powder + lime + oil cakes.

Used neem cake, muriate of potash, Blitox-50, Bavistin 0.1 per cent, marigold leaf extract 5 per cent, garlic bulb extract 5 per cent and neem leaf extract 5 per cent alone and in combination treatments against wilt complex fungi (*R. solani*, *F. oxysporum*, *Sclerotium rolfsii* and *Macrophomina phaseolina*) in lentil and inferred that application of neem cake (250 kg/ha), neem leaf extract (5%) and seed treatment with Bavistin (1 g/kg seed) followed by drenching Blitox-50 (0.03%) reduced the incidence of wilt complex [32]. Integrated different management practices including fumigation, biocontrol agents and seed treatment with fungicides against *Pythium aphanidermatum* causing damping-off in tomato [33] and observed that nursery bed treatment with formalin, basamid and their combination with seed treatment (captaf 0.3%) provided 100 per cent control of the disease, whereas, nursery bed treatment with carbendazim granules and *Trichoderma viride* was slightly less effective (4% disease incidence).

Reported that damping-off (*Fusarium* and *Pythium*) of egg plant was lowest when soil was drenched with formaldehyde (5 %) combined with seed treatment with thiram @ 3 g/kg seed and drenching of copper oxychloride @ 0.3 per cent after seedling emergence [16]. Observed that soil disinfection with lime one month before seedling transplantation and the use of *Pseudomonas fluorescens* as biocontrol agent effectively minimized the bacterial wilt in tomato caused by *Ralstonia solanacearum* [34]. Reported effectiveness of soil solarization in combination with *P. fluorescens* and *Trichoderma harzianum* against *Pythium* spp. causing damping-off in tomato [35].

References

1. Dreschler, E. 1926. The cottony leak of cucumber caused by *Pythium aphanidermatum*. *Journal of Agricultural Research*, 30: 1035-1042.

2. Gattani, M. L. and Koul, T. N. 1951. Damping off of tomato seedlings, its causes and control. *Indian Phytopathology*, 4: 156-161.
3. Hooker, A. L. 1953. Relative pathogenicity of *Pythium* species attacking seedlings of corn. *Iowa Academy Sciences*, 60: 163-166.
4. Kilpatrick, R. A. 1968. Seedlings reaction of barley, oats and wheat to *Pythium* species. *Plant Disease Reporter*, 52: 209-212.
5. Waterhouse, G. M. 1968. *The genus Pythium*. Mycological Papers. Vol. 110, pp 1-71 Commonwealth Mycological Institute, Kew, London, U.K.
6. Grover, R. K. and Dutt, S. 1973. Morphological and pathological variability in *Pythium aphanidermatum*. *Indian Phytopathology*, 26: 237-244.
7. Rangaswami, G. and Mahadevan, A. 2006. *Diseases of Crop Plants in India*. pp.534. Prentice Hall of India Private limited, New Delhi, India.
8. Ogoshi, A. and Ui, T. 1983. Diversity of clones within an anastomosis group of *Rhizoctonia solani* Kuhn in a field. *Annual Review of the Japan Phytopathological Society*, 49: 239-245.
9. Alavi, A., Saber, M. and Akhavizadigam, M. D. 1986. Rhizoctonia crown and root rot of pepper in Iran. *Iranian Journal of Plant Pathology*, 22: 9-12.
10. Mushtaq, M. and Hashmi, M. H. 1997. Fungi associated with wilt disease of capsicum in Sindh Pakistan. *Pakistan Journal of Botany*, 29: 217-222.
11. Zapta, R. L., Palmucci, H. E., Murray, V. B. and Lapez, M. V. (2001a). Biological trials to control damping off in brinjal with fluorescent *Pseudomonas* and *Trichoderma harzianum*. *Revist-de-Faculta-de-Agronomia-Universidad-de-Buenio-Aires*, 21: 207-211.
12. Pandey A.K., Rajak, R.C. Hasija, S.K. (2001). Biotechnological development of ecofriendly mycoherbicides Innovative Approaches in Microbiology 1-21.
13. Zapta *et al.* (2001). Grapevine culture in trenches: root growth and dry matter partitioning. *Australian Journal of Grape and Wine Research*, 7: 127-131.
14. Herrado, U. M. T., Garcia, G. V. M. and Marquina, J. T. 2004. Fusarium wilt on eggplant in Almeria (Spain). *Bolentin-de-Sanidad-Vegetal-Plagas*, 30: 85-92.
15. Altinok, H. H. 2005. First report of Fusarium wilt of eggplant by *Fusarium oxysporum* f. sp. *melanogenae* in Turkey. *Plant Pathology*, 54: 577.
16. Datar, V. V. 2007. Investigations of pre- and post-emergence mortality in eggplant (*Solanum melongena* L.). *Indian Phytopathology*, 60: 156-161.
17. Sally, A., Miller, R. C. R. and Richard, M. R. 2007. Fusarium and Verticillium wilts of tomato, potato, pepper and eggplant. *Extension Factsheet*. pp. 1-3. Ohio State University, U.S.A.
18. Siva, N., Ganesan, S., Banumathy, N. and Muthuchelian. 2008. Antifungal effect of leaf extracts of some medicinal plants against *Fusarium oxysporum* causing wilt disease of *Solanum melongena* L. *Enthobotanicals Leaflets*, 12: 156-163.
19. Kelman, A. 1953. *The bacterial wilt caused by Pseudomonas solanacearum, A Literature Review and Bibliography*. Technical Bulletin No.99. p 194. North Carolina Experiment Station.
20. Doolittle, S. P. 1953. Diseases of pepper. In: *Year Book of Agriculture* pp. 465-469. Oxford and IBH Publishing Company, New Delhi.
21. Das, C. R. and Chattopadhyay, S. B. 1955. Bacterial wilt of eggplants. *Indian Phytopathology*, 8: 130-135.
22. Kelman, A. 1954. A relationship of pathogenicity of *Pseudomonas solanacearum* to colony appearance in a T. T. C. medium. *Phytopathology*, 44: 693-695.
23. Hayward, A. C. 1960. A method of characterizing *Pseudomonas solanacearum*. *Nature*, 186: 405-406.
24. Jyothi, A. R., Koshy, A., Mathew, J. and Abraham, K. 1993. Bacterial wilt of chilies in Kerala and reaction of certain chilli accessions to *Pseudomonas solanacearum*. *Bacterial Wilt Newsletter*, 9: 6.
25. Chatterjee, S., Mukherjee, N. and Khatua, D. C. 1997. Status of bacterial wilt disease in West Bengal. *Journal of Interacademica*, 1: 97-99.
26. Bhardwaj, L. N., Shyam, K. R. and Thakur, P. D. 1987. Effect of seed and soil treatments on the incidence of damping-off and black rot in cauliflower seedlings. *Pesticides*, 21: 13-16.
27. Sawant, I. S. and Mukhopadhyay, A. N. 1990. Integration of metalaxyl with *Trichoderma harzianum* for the control of Pythium damping off in sugarbeet. *Indian Phytopathology*, 43: 535-541.
28. Khulbe, D. and Chaube, H. S. 2002. Effect of soil solarization and its integration with bioagents and fungicides on the incidence of damping off and plant growth response of vegetable crops in nursery. *Journal of Mycology and Plant Pathology*, 32: 412.
29. Ramesh, R. and Manjunath, B. L. 2002. Effect of recycled manurial resources, biocontrol agents and fungicides on the incidence of brinjal wilt. *Resources Management in Plant Protection during 21st century*, Hyderabad, India 2: 15-18.
30. De, R. K., Dwivedi, R. P. and Narain, U. 2003. Biological control of lentil wilt caused by *Fusarium oxysporum* f. sp. *lentis*. *Annals of Plant Protection Sciences*, 11: 46-52.

31. Nath, R., Dutta, S., Laha, S.K. (2004). *Pseudomonas fluorescens* - A potential biocontrol weapon to combat bacterial wilt of tomato. *Journal of Mycology and Plant Pathology*, 34(3):974.
32. Sinha, R. K. P. and Sinha, B. B. P. 2004. Effect of potash, botanicals and fungicides against wilt disease complex in lentil. *Annals of Plant Protection Sciences*, 12: 454-455.
33. Sharma, H. R. and Gupta, S. K. 2004. Management of damping off of tomato seedlings through integrated management practices. *Journal of Mycology and Plant Pathology*, 34: 982.
34. Biswas, S. and Singh, N. P. 2008. Integrated management of wilt of tomato caused by *Ralstonia solanacearum*. *Journal of Mycology and Plant Pathology*, 38: 18-20.
35. Jayaraj, J. and Radhakrishnan, N. V. 2008. Enhanced activity of introduced biocontrol agents in solarized soils and its implications on the integrated control of tomato damping off caused by *Pythium* spp. *Plant Soil*, 304: 189-197.